Determination of Biological Properties of Chlorella vulgaris C1 Extracts

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Abstract

In this study, anti-genotoxic and anti-oxidant activities of algal type Chlorella vulgaris C1 were investigated. Dried samples in the Soxhlet device; It was extracted with acetone, methanol, distilled methanol and dimethyl sulfoxide. The anti-oxidant and anti-mutagenic properties of the extracts were determined by micronucleus and Trolox equivalent anti-oxidant capacity methods. It was observed that the extracts investigated showed anti-genotoxic properties by preventing the mutagenic activity of AFB₁ in the tested concentrations. In addition, methanol and acetone extracts were found to show relatively higher anti-oxidant and anti-mutagenic activity compared to other extracts. The results show that these natural algae have the ability to reduce the mutagenic activity of AFB₁, suggesting that antioxidant activities may play a role in anti-genotoxic activity mechanisms. Due to these properties, the use of Chlorella vulgaris C1 in the pharmaceutical and food industry can be expanded.

Keywords: Aflatoxin B₁, *Chlorella vulgaris* C1, Micronuclei, Oxidative.

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1. Introduction

chlorophyll-containing Algae are simple organisms composed of one cell or cells grouped together in colonies (Wynne and Bold 1985). They are consisting of proteins, lipids, essential amino acids, anti-oxidant pigments, fatty acids, vitamins, carotenoids and other bioactive compounds that express unique features for the development of pharmaceuticals, nutraceuticals, cosmetics, biofuel industry and CO₂ fixation ability (Jin et al. 2006; Oncel 2013; Pulz and Gross 2004). Chlorella vulgaris (CV) is a microscopic single-celled green alga. It is known that microalgal phyla provides pharmacological and chemical diversity innovation. In addition, microalgae are considered to be important producers of some bioactive compounds that are highly found in marine resources (Shimizu 1996).

Previous studies have been reported that CV has rich source of anti-oxidants, such as carotene and tocopherol, lutein and ascorbic acid, also supplying large quantities of vitamins, minerals, dietary fiber and essential fatty acids (Rodriguez-Garcia and Guil-Guerrero 2008). In additional, some researchers showed that treatment with CV has protective activities against stress, tumors, infections and high-fat-diet-induced insulin resistance (Dantas and Queiroz 1999; Ramos et al. 2010). Therefore, Aremu et al. (2016) reported that phytochemical content and biological activities of the *Chlorella* strains were affected by strain, harvest time and N levels.

Negishi et al. (1991) demonstrated that a chlorophyll sample prepared from CV has anti-

genotoxic activity using *Salmonella* and wing spot *Drosophila* test systems. There have been many studies reporting related to especially anti-oxidant activity utilization of CV. This organism showed a shielder effect against heavy metals and other harmful components (such as lead, cadmium, and naphthalene) by decreasing remarkably the oxidative stress reduced by these detrimental compounds, and increasing the anti-oxidant activity in the organisms of tested animals (Shim et al. 2008; Yun et al. 2011).

Recently, there has been an increase in naturally occurring anti-oxidants research for their use in food or medicinal compounds to replace synthetic anti-oxidants, which are being restricted due to their carcinogenicity (Velioglu et al. 1998).

Research and determination of anti-mutagenic and anti-oxidant activities of these compounds has become an important strategy in the treatments of many human diseases related to mutations (Celikler et al. 2009; Okai et al. 1996).

In addition, owing to the changefulness in chemical and aromatic compounds, seaweeds are conventionally the cosmetic used in pharmaceutical industries as culinary herbs. They have also been used as a traditional food and medicine for healing helminths infections, eczema and gout, most especially by people in coastal areas of several countries (Hoppe 1979). Especially, some metabolites from these seaweeds act as protective compounds against endogenic and exogenic agents threatening the genome. There are several studies conducted to designate the biological activity of these including immunostimulant, metabolites microbial, anti-oxidative, anti-ulcerogenic, antiinflammatory, analgesic/antipyretic, anti-tumor and anti-mutagenicity activity assays (Khan et al. 1988; Leitte-Silva et al. 2007).

Therefore, in this study, we investigated genotoxic and anti-genotoxic properties of the extracts of *C. vulgaris* C1 by using micronuclei (MN) assay. In addition, the total anti-oxidant activity was measured in order to clarify the mechanism underlying the anti-genotoxic effects of CV. It is also the first study to determine the mutagenic and anti-mutagenic activity depending on the biological activity of *C. vulgaris* C1 isolated and identified from Mogan Lake (Turkey-Ankara). On the other hand, the mutagenic and anti-mutagenic effects of different extracts obtained from the same strain were also

evaluated in terms of the antagonistic and synergistic effects of the extracts.

2. Materials and Methods

2.1. Culturing and growth conditions

The seaweeds were collected from Mogan Lake in Ankara, Turkey. Micromanipulation technique was used to isolate CV from mix culture and the axenic cultures were obtained. Collection and isolation of microalgae were made in compliance with Rippka (1988) and Attalah et al. (2019).

2.2. Preparation of the extracts

The extracts were prepared according to the methods of Khan et al. (1988) and Vlaschos et al. (1996). The dried extracts were resuspended in 3 ml of each solvent and preserved at 4 °C for further use in different bioassays (Deshmukh and Puranik 2010; Prakash et al. 2011).

2.3. Quantitative analysis of chemical constituents

TOF-LC / MS was used for quantitative analysis and was performed at Çankırı Karatekin University (Erenler et al. 2015).

2.4. Micronuclei tests

For the micronucleus test, Nartop et al. (2020) determined procedure was applied. Experiments on different extracts of Chlorella vulgaris C1 (10, 20 and 40 μ g/ml) were carried out with 6 groups as follows:

Culture 1: Control

Culture 2: AFB1 (5 µM)

Culture 3: CVE (20 µg/ml)

Culture 4: AFB1 $(5 \mu M)$ + CVE $(10 \mu g/ml)$

Culture 5: AFB1 (5 μ M) + CVE (20 μ g/ml)

Culture 6: AFB1 $(5 \mu M) + CVE (40 \mu g/ml)$

2.5. Total antioxidant status

Total anti-oxidant activity was measured by Trolox equivalent anti-oxidant capacity (TAC) assay. The assay is calibrated with a stable anti-oxidant standard solution called Trolox Equivalent, that is a vitamin E analogue (Yıldırım et al. 2013). The automated TAS experiment was carried out by commercially available kits (Total Antioxidant Status, Rel Assay Diagnostics, Turkey; Turkez et al. (2010). Regents from the kits were added on to the samples from the extracts.

2.6. Statistical analysis

Data were analyzed and treatments compared using the one-way ANOVA with 95% confidence limits (p<0.05), according to Duncan's multiple range tests (SPSS 15.0 Version).

3. Results

The seaweed extracts showed anti-oxidant activity to various degrees (Tab. 1). As shown in Tab. 1, acetone and methanol extracts of CV exhibited high anti-oxidant activity which was significantly different compared with other CV extracts.

MN frequencies of the experimental groups are given in Tab. 2. MN frequency in AFB1 treated group (5 μM) was higher than that in the control group (p <0.05). There was a significant decrease in the MN frequency in CV extracts - treated group when compared with the groups receiving AFB₁. In additional, the composition of CV extracts and the relative amounts of the components are analysis as shown in the Tab. 3. According to these results, salicylic acid (0.893 mg / kg) was found higher in methanol extract while vanillic acid (2.107 mg / kg) higher in acetone extraction. It is also seen that vanillic acid (0.714 mg / kg) compounds in dimethyl sulfoxide (DMSO) extract are higher than other compounds. This result shows that alone the activity of anti-oxidant of methanol extract was the same DMSO + acetone mixture (4.14 mmol / L) (4.37 mmol/L).

Table 1. Total antioxidant status of Chlorella vulgaris C1 extracts

Algal species	Extract	Abbreviation	Antioxidant (mmol/L)
C. vulgaris C1	DMSO+	CVDA	4.37
	Acetone		
	DMSO	CVD	0.23
	DMSO+	CVDM	0.92
	Methanol		
	Distillate	CVdM	0.23
	Methanol		
	Methanol	CVM	4.14

Table 2. The effects of AFB1 and extracts of Chlorella vulgaris C1 (CVE) on MN

SE AFB₁ + CVE AFB₁ + CVE AFB₁ + CVE Control AFB_1 was used as positive controls for human blood cells. Values of MN (a, b, c, d) are significantly different compared to negative control (P < 0.05) Test Items 5 μM + 40 μM $5 \mu M + 10 \mu M$ 20 µM $5 \mu M + 20 \mu M$ Concentrations $4.72 \pm 0.48^{\circ}$ **MN** numbers 2.72 ± 0.19^{a} 2.96 ± 0.27^{ac} $3.21 \pm 0.36^{\circ}$ 2.56 ± 0.13 2.45 ± 0.01^{a} ± S.E (CVDA 4.72 ± 0.48^{d} 2.45 ± 0.01^{a} 2.52 ± 0.13^{a} $4.35 \pm 0.36^{\circ}$ 3.98± 0.19bc 4.14 ± 0.27^{co} **MN** numbers ± S.E (CVD) $3.89 \pm 0.36^{\circ}$ MN numbers ± S.E (CVDM) 3.70 ± 0.27 4.72 ± 0.48^{d} 2.45 ± 0.01^{a} 3.61 ± 0.19^{cb} 2.63 ± 0.13^{a} **MN** numbers ± S.E (CVdM) 3.92 ± 0.19^{cb} 4.03 ± 0.27 4.28 ± 0.36^{cc} 2.64 ± 0.13^{a} $4.72 \pm 0.48^{\circ}$ 2.45 ± 0.01^{a} 4.90 ± 0.43^{a} $5.27 \pm 0.08^{\circ}$ $5.58 \pm 0.31^{\circ}$ 4.82± 0.22^a 6.72 ± 0.48^{d} 4.45 ± 0.01^{a} ± S.E (CVM) MN numbers

Table 3. Chemical composition of C. vulgaris C1 with different extracts. (mg/kg)

Compound	DMSO	Methanol	Acetone
Gallic acid	0.048	-	-
Gentisic acid	0.1488	-	-
Caftaric acid	-	0.0341	-
Chlorogenic	-	-	-
acid			
Catechin	1.84	-	-
P-	1.9247	0.6028	0.8824
hydroxybenzoic			
acid			
Protocatechuic	-	-	-
acid			
Caffeic acid	0.1734	0.5572	0.5209
Rutin	0.7168	-	-
p-coumaric	0.067	0.02486	0.6318
acid			
Chicoric acid	-	-	-
Ferulic acid	0.0471	0.3646	-
Hesperidin	-	-	-
Apigenin-7-	-	-	-
glucoside			
Rosmarinic acid	0.0023	-	-
Protocatechuic	0.5312	0.562	0.962
acid ethyl ester			
Salicylic acid	0.498	0.893	0.163
Quercetin	-	-	-
Cinnamic acid	-	-	-
Naringenin	-	-	-
Kaempferol	-	0.037	-
Vanillic acid	0.714	0.0374	2.107
Caffeic acid	0.1734	0.5572	0.5209
Rutin	0.7168	-	-
p-coumaric	0.067	0.02486	0.6318
acid			

4. Discussion

CV is known as a functional food source. Therefore, in this study, the antagonistic effects of extracts of CV were studied against AFB_1 mutation

agents in the peripheral blood lymphocytes using MN test systems. This agent is known to stimulate the release of free radicals, including reactive oxygen species that cause chromosomal aberrations (Ceker et al. 2018; Orhan et al. 2016). CVE showed great antimutagenic potential against AFB₁. This antimutagenic activity may be explained with inhibitor activities of the CVE on the formation of free radicals. In order to elucidate the anti-mutagenic activities of CVE, the amount of anti-oxidants was determined. Our results showed anti-oxidant activity to various degrees all extracts.

As shown in Tab. 2 methanol and acetone extract exhibited relatively high anti-oxidant activity. This is the first scientific report on the anti-genotoxic and protective potential of CVE.

Recent studies have reported that several algal extracts have anti-cancer, immunoregulator, immunostimulant, anti-oxidant, anti-microbial activities and strong anti-genotoxic activity in human lymphocytes in vitro (Celikler et al., 2008; Faulkner 2000; Koyanagı et al. 2003; Leitte-Silva et al. 2007; Okai et al. 1996; Varella et al. 2004; Yamamoto et al. 1986).

Several authors have reported that algal extracts have anti-oxidant and anti-mutagenic/anti-carcinogenic activities due to compounds, such as β -carotene, lutein and chlorophyll-related derivatives isolated from this species (Benedetti et al. 2004; Kotake-Nara et al. 2001).

The phenolic compounds of CV such as vanillic acid, caffeic acid, ferulic acid, protocatechuic acid, pcoumaric acid, 4-hydroxybenzoic acid, salicylic acid, caftaric acid, camphenol have been determined in our previous unpublished studies. Among the and of CV vanillic, fumaric, contents caffeic, protocatechuic and caftaric acid have been reported that have to anti-mutagenic activities in previous studies and these effects could be related to its antioxidant potential. Other phenolic compounds have determined anti-oxidant activities but there have not been sufficiently studies related to anti-mutagenic potential (Chen et al. 2016; Safaeian et al. 2016; Zhang et al. 2011).

In addition, the results obtained in this study suggest that to understand the anti-oxidant activities of extracts obtained from CV may not suffice compounds derived from a single solvent. It has been demonstrated that synergistic and antagonistic effects

of the compounds obtained from these solvents may also be present. Anti-mutagenic potentials and changes were also investigated with these interactions.

The algal extracts examined in our study showed a great anti-oxidant property. According to the results obtained from MN test system, CV has a significant anti-genotoxic effect. Taken together, the results of our study indicate that anti-genotoxic effect of CV could be related to its anti-oxidant potential. Although the performed test systems showed important data including anti-genotoxic and anti-oxidant potential of the CV, further studies are needed.

The clarification of each of the contents of CV can extend the usage of it in treating some diseases. These compounds are valuable towards an extension of the use of drugs as new phytotherapeutic or preservative ingredients, besides their consolidated ethno medical use. In addition, CV and compounds can be use in chemotherapeutic drugs with the purpose of more effective curation with least toxicity.

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References

- AREMU, A. O., MASONDO, N. A., MOLNÁR, Z., STIRK, W. A., ÖRDÖG, V., & VAN STADEN, J. (2016), Changes in phytochemical content and pharmacological activities of three Chlorella strains grown in different nitrogen conditions. Journal of applied phycology, 28(1), 149-159.
- ATTALAH, S., WALLER, P., STEICHEN, S., GAO, S., BROWN, C., OGDEN, K., & BROWN, J. K. (2019), Application of deoxygenation-aeration cycling to control the predatory bacterium Vampirovibrio chlorellavorus in Chlorella sorokiniana cultures. Algal Research, 39, 101427.
- BENEDETTI, S., BENVENUTI, F., PAGLIARANI, S., FRANCOGLI, S., SCOGLIO, S., & CANESTRARI, F. (2004), Antioxidant properties of a novel phycocyanin extract from the blue-green alga Aphanizomenon flosaquae. Life sciences, 75(19), 2353-2362.

- CEKER, S., ORHAN, F., SEZEN, S., GULLUCE, M., OZKAN, H., ASLAN, A., & AGAR, G. (2018), Anti-mutagenic and anti-oxidant potencies of Cetraria aculeata (Schreb.) Fr., Cladonia chlorophaea (Flörke ex sommerf.) spreng. and Cetrelia olivetorum (Nyl.) WL Culb. & CF Culb. Iranian journal of pharmaceutical research: IJPR, 17(1), 326.
- CELIKLER, S., VATAN, O., YILDIZ, G., & BILALOGLU, R. (2009), Evaluation of antioxidative, genotoxic and antigenotoxic potency of Codium tomentosum Stackhouse ethanolic extract in human lymphocytes in vitro. Food and chemical toxicology, 47(4), 796-801.
- CELIKLER, S., YILDIZ, G., VATAN, O., & BILALOGLU, R. (2008), In vitro antigenotoxicity of Ulva rigida C. Agardh (Chlorophyceae) extract against induction of chromosome aberration, sister chromatid exchange and micronuclei by mutagenic agent MMC. Biomedical and Environmental Sciences, 21(6), 492-498.
- CHEN, H., ZHOU, Y., SHAO, Y., & CHEN, F. (2016), Free phenolic acids in Shanxi aged vinegar: Changes during aging and synergistic antioxidant activities. International journal of food properties, 19(6), 1183-1193.
- COUNTRYMAN, P. I., & HEDDLE, J. A. (1976), The production of micronuclei from chromosome aberrations in irradiated cultures of human lymphocytes. Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis, 41(2-3), 321-331.
- DANTAS, D. C., & QUEIROZ, M. L. (1999), Effects of Chlorella vulgaris on bone marrow progenitor cells of mice infected with Listeria monocytogenes. International journal of immunopharmacology, 21(8), 499-508.
- DEVENDRA, V. D. & P., R. PURANIK. (2010), Application of Plackett-Burman design to evaluate media components affecting antibacterial activity of alkaliphilic cyanobacteria isolated from Lonar Lake. Türk Biyokimya Dergisi [Turkish Journal of Biochemistry—Turk J Biochem], 35(2), 114-120.
- ERENLER, R., TELCI, I., ULUTAS, M., DEMIRTAS, I., GUL, F., ELMASTAS, M., & KAYIR, O. (2015), Chemical Constituents, Quantitative Analysis and Antioxidant Activities of E chinacea purpurea (L.) M oench and E chinacea pallida (N utt.) N utt. Journal of Food Biochemistry, 39(5), 622-630.

- FAULKNER, D. J. (2000), Highlights of marine natural products chemistry (1972–1999). Natural product reports, 17(1), 1-6.
- HOPPE, H. A. (1979), Marine algae and their products and constituents in pharmacy. Marine algae in pharmaceutical science, editors, Heinz A. Hoppe, Tore Levring, Yukio Tanaka.
- JIN, H.-F., LIM, B.-R., & LEE, K. (2006), Influence of nitrate feeding on carbon dioxide fixation by microalgae. Journal of Environmental Science and Health Part A, 41(12), 2813-2824.
- KHAN, N., RAHMAN, M., & NUR-E-KAMAL, M. (1988), Antibacterial activity of Euphorbia thymifolia Linn.
- KOTAKE-NARA, E., KUSHIRO, M., ZHANG, H., SUGAWARA, T., MIYASHITA, K., & NAGAO, A. (2001), Carotenoids affect proliferation of human prostate cancer cells. The Journal of nutrition, 131(12), 3303-3306.
- KOYANAGI, S., TANIGAWA, N., NAKAGAWA, H., SOEDA, S., & SHIMENO, H. (2003), Oversulfation of fucoidan enhances its antiangiogenic and antitumor activities. Biochemical pharmacology, 65(2), 173-179.
- KUSANO, C., & FERRARI, B. (2008), Total antioxidant capacity: a biomarker in biomedical and nutritional studies. J Cell Mol Biol, 7(1), 1-15.
- LEITE-SILVA, C., GUSMÃO, C. L. S., & TAKAHASHI, C. S. (2007), Genotoxic and antigenotoxic effects of Fucus vesiculosus extract on cultured human lymphocytes using the chromosome aberration and Comet assays. Genetics and Molecular Biology, 30(1), 105-111.
- NEGISHI, T., SHIOTANI, T., FUJIKAWA, K., & HAYATSU, H. (1991), The genotoxicities of N-nitrosamines in Drosophila melanogaster in vivo: the correlation of mutagenicity in the wing spot test with the DNA damages detected by the DNA-repair test. Mutation Research/Environmental Mutagenesis and Related Subjects, 252(2), 119-128.
- OKAI, Y., HIGASHI-OKAI, K., YANO, Y., & OTANI, S. (1996), Identification of antimutagenic substances in an extract of edible red alga, Porphyra tenera (Asadusanori). Cancer letters, 100(1-2), 235-240.
- ONCEL, S. S. (2013), Microalgae for a macroenergy world. Renewable and Sustainable Energy Reviews, 26, 241-264.
- ORHAN, F., CEKER, S., ANAR, M., AGAR, G., ARASOGLU, T., & GULLUCE, M. (2016), Protective effects of three luteolin derivatives on aflatoxin B 1-induced genotoxicity on

- human blood cells. Medicinal chemistry research, 25(11), 2567-2577.
- PRAKASH, J. W., MARIMUTHU, J., & JEEVA, S. (2011), Antimicrobial activity of certain fresh water microalgae from Thamirabarani River, Tamil Nadu, South India. Asian Pacific Journal of Tropical Biomedicine, 1(2), S170-S173.
- PULZ, O., & GROSS, W. (2004), Valuable products from biotechnology of microalgae. Applied microbiology and biotechnology, 65(6), 635-648
- RAMOS, A. L., TORELLO, C. O., & QUEIROZ, M. L. (2010), Chlorella vulgaris modulates immunomyelopoietic activity and enhances the resistance of tumor-bearing mice. Nutrition and cancer, 62(8), 1170-1180.
- RIPPKA, R. (1988), [1] Isolation and purification of cyanobacteria. In Methods in enzymology (Vol. 167, pp. 3-27): Elsevier.
- RODRIGUEZ-GARCIA, I., & GUIL-GUERRERO, J. L. (2008), Evaluation of the antioxidant activity of three microalgal species for use as dietary supplements and in the preservation of foods. Food chemistry, 108(3), 1023-1026.
- SAFAEIAN, L., HAJHASHEMI, V., JAVANMARD, S. H., & NADERI, H. S. (2016), The effect of protocatechuic acid on blood pressure and oxidative stress in glucocorticoid-induced hypertension in rat. Iranian journal of pharmaceutical research: IJPR, 15(Suppl), 83.
- SHIM, J.-Y., SHIN, H.-S., HAN, J.-G., PARK, H.-S., LIM, B.-L., CHUNG, K.-W., & OM, A.-S. (2008), Protective effects of Chlorella vulgaris on liver toxicity in cadmium-administered rats. Journal of medicinal food, 11(3), 479-485.
- SHIMIZU, Y. (1996), Microalgal metabolites: a new perspective. Annual review of microbiology, 50(1), 431-465.
- TURKEZ, H., GEYIKOGLU, F., ASLAN, A., KARAGOZ, Y., TURKEZ, O., & ANAR, M. (2010), Antimutagenic effects of lichen Pseudovernia furfuracea (L.) Zoph. extracts against the mutagenicity of aflatoxin B1 in vitro. Toxicology and Industrial Health, 26(9), 625-631.
- VARELLA, S. D., POZETTI, G. L., VILEGAS, W., & VARANDA, E. A. (2004), Mutagenic activity in waste from an aluminum products factory in Salmonella/microsome assay. Toxicology in vitro, 18(6), 895-900.
- VELIOGLU, Y., MAZZA, G., GAO, L., & OOMAH, B. (1998), Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. Journal of agricultural and food chemistry, 46(10), 4113-4117.

- VLACHOS, V., CRITCHLEY, A., & VON HOLY, A. (1996), Establishment of a protocol for testing antimicrobial activity in southern African macroalgae. Microbios, 88(355), 115-123.
- WYNNE, M. J., & BOLD, H. (1985), Introduction to the Algae: Structure and Reproduction: Prentice-Hall, Incorporated.
- YAMAMOTO, I., MARUYAMA, H., TAKAHASHI, M., & KOMIYAMA, K. (1986), The effect of dietary or intraperitoneally injected seaweed preparations on the growth of sarcoma-180 cells subcutaneously implanted into mice. Cancer letters, 30(2), 125-131.
- YILDIRIM, N. C., PAKSOY, M. Y., YUCE, E., & YILDIRIM, N. (2013), Total antioxidant status and antifungal activities of endemic geophytic plants collected from Munzur Valley in Tunceli, Turkey. Digest Journal of Nanomaterials & Biostructures (DJNB), 8(1), 403-408.
- YUN, H., KIM, I., KWON, S.-H., KANG, J.-S., & OM, A.-S. (2011), Protective effect of Chlorella vulgaris against lead-induced oxidative stress in rat brains. Journal of Health Science, 57(3), 245-254.
- ZHANG, X., ISHIDA, R., YUHARA, Y., KAMIYA, T., HATANO, T., OKAMOTO, G., & ARIMOTO-KOBAYASHI, S. (2011), Antigenotoxic activity of Vitis coignetiae Pulliat towards heterocyclic amines and isolation and identification of caftaric acid as an antimutagenic component from the juice. Mutation Research/Genetic Toxicology and Environmental Mutagenesis, 723(2), 182-189.